

Coding and classification of D-galactose, N-acetyl-D-galactosamine, and β -D-Galp-[1 $\rightarrow$ 3(4)]- β -D-GlcNAc, specificities of applied lectins^{*,†}

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ABSTRACT

Grouping of lectin-binding properties, based on determinant structure rather than monosaccharide-inhibition pattern, should facilitate the selection of lectins as structural probes for glycans, as well as for the interpretation of the distribution and the properties of the carbohydrate chains on the cell surface. Based on the binding specificities studied with glycan by precipitin-inhibition, competitive-binding, and hemagglutinin-inhibition assays, twenty D-galactose- or N-acetyl-D-galactosamine- (or both)-specific lectins have been divided into six classes according to their specificity for a disaccharide unit, as all or part of the determinants, and the α -D-GalpNAc-(1 $\rightarrow$ 3)-Ser(Thr) unit of the glycopeptide chain. A scheme of classification is shown as follows: (a) F-specific lectins [α -D-GalpNAc-(1 $\rightarrow$ 3)-D-GalNAc, Forssman specific disaccharide]: *Dolichos biflorus* (DBL), *Helix pomatia* (HPL), hog peanut (ABL, *Amphicarpaea bracteata*), and *Wistaria floribunda* (WFL) lectins. (b) A-specific lectins [α -D-GalpNAc-(1 $\rightarrow$ 3)-D-Gal blood group A-specific disaccharide]: *Griffonia* (*Bandeiraea*) *simplicifolia*-A₄ (GSI-A₄), lima bean (LBL), soy bean (SBL), *Vicia villosa* (VVL), *Wistaria floribunda* (WFL), *Dolichos biflorus* (DBL), and *Helix pomatia* (HPL) lectins. (c) Tn-specific lectins [α -D-GalpNAc-(1 $\rightarrow$ 3)-Ser(Thr) of the protein core]: *Vicia villosa* B₄ (VVL-B₄), *Salvia sclarea* (SSL), *Maclura pomifera* (MPL), *Bauhinia purpurea alba* (BPL), HPL, and WFL, lectins. (d) T-specific lectins [β -D-Galp-(1 $\rightarrow$ 3)-D-GalNAc, the mucin-type sugar sequences on human erythrocyte membrane and T antigen, or the terminal, nonreducing disaccharide end-groups of the gangliosides]: Peanut (PNA), *Bauhinia purpurea alba* (BPL), *Maclura pomifera* (MPL), *Sophora japonica* (SJL), *Artocarpus integrifolia* (Jacalin, AIL), and *Artocarpus lakoocha* (Artocarpin) lectins. (e) Type I and II specific lectins [β -D-Galp-(1 $\rightarrow$ 3 or 4)-D-GlcNAc, the disaccharide residues at the nonreducing end of the carbohydrate chains derived from either N- or O-glycans]: *Ricinus communis* agglutinin (RCA1), *Datura stramonium* (TAL, Thorn apple), *Erythrina cristagalli* (ECL, Coral tree), and *Geodia cydonium* (GCL), lectins. (f) B-specific lectin [α -D-Galp-(1 $\rightarrow$ 3)- β -D-Galp, human blood group B-specific disaccharide]: *Griffonia* (*Bandeiraea*) *simplicifolia* B₄ (GSI-B₄) lectin. Many other GalNAc- or Gal- (or both)-specific lectins that can be used as tools are also described.

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[†] This paper is condensed from "Differential Binding Properties of GalNAc and/or Gal Specific Lectins" and "A Guide for Carbohydrate Specificities of Lectins" in "The Molecular Immunology of Complex Carbohydrates", A.M. Wu (Ed.), *Adv. Exp. Med. Biol.*, 228 (1988) 205–263 and 822–832, Plenum, New York. The lectins reported in this paper are those that can be used as tools.

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INTRODUCTION

Lectins have been widely used to study the structural and functional role of cell surface carbohydrates¹⁻⁶, to detect sugar components on normal and neoplastic cell surfaces¹, to isolate mutants resistant to the cytotoxic action of some lectins⁷⁻¹⁰, and to isolate and characterize glycoconjugates^{5,11-14}. They require configurational and structural complementarity of sugars for interaction to occur. All lectin molecules have more than two carbohydrate-binding sites, a property resulting in their ability to agglutinate cells or to precipitate complex carbohydrates^{1,4,15,16}. By the early eighties, the carbohydrate specificities of many lectins were grouped by the ability of monosaccharides or their glycosides to inhibit lectin-induced hemagglutination^{2,15,17,18}. Even lectins of the same apparent monosaccharide specificity were found to demonstrate different reactivities toward different oligosaccharide chains, and differential affinities to animal cells and glycoproteins, which implies that they have their own binding specificity extending beyond the monosaccharide unit^{1-4,15,18}. Today, the concept of the combining sites of many lectins extending to five or more sugars has been accepted, and the fact that multibranched oligosaccharides exhibit a significant increment in their lectin-binding reactivities as compared with the linear counterparts has also been recognized. For example, *Datura stramonium* (TAL, Thorn apple) lectin shows a specificity for a biantennary pentasaccharide, which contains two branches, β -D-Galp-(1 $\rightarrow$ 4)- β -D-GlcNAc-(1 $\rightarrow$ 6)-D-Man and β -D-Galp-(1 $\rightarrow$ 4)- β -D-GlcNAc-(1 $\rightarrow$ 2)-D-Man as nonreducing end-groups, 480-fold higher than for that of its branch trisaccharides¹⁹. Furthermore, some lectins having broad spectra of binding properties possess dual or multiple affinities to various disaccharides. Such lectins as *Wistaria floribunda* is specific for α -D-GalpNAc-(1 $\rightarrow$ 3)-D-GalNAc, α -D-GalpNAc-(1 $\rightarrow$ 3)-D-Gal, α -D-GalpNAc-(1 $\rightarrow$ 3)-Ser(Thr), and β -D-Gal-(1 $\rightarrow$ 4 or 3)-D-GlcNAc units; *Bauhinia purpurea alba* for β -D-Gal-(1 $\rightarrow$ 3)-D-GalNAc, α -D-GalpNAc-(1 $\rightarrow$ 3)-Ser(Thr), and β -D-Galp-(1 $\rightarrow$ 4 or 3)-D-GlcNAc units, and *Datura stramonium* for N-acetyllactosamine and N-acetylchitobiose units^{2,19}. Thus, organizing and grouping binding properties of lectins, based on disaccharide determinants and α -D-GalpNAc-(1 $\rightarrow$ 3)-Ser(Thr) sequence, rather than monosaccharide-inhibition pattern, should facilitate the selection of lectins as structural probes for glycans, as well as the interpretation of the distribution and properties of the carbohydrate chains on the cell surface. From the information provided by inhibition assays (Tables I and II)*, the lectins selected in this paper were divided into six classes according to their specificity for the α -D-GalpNAc-(1 $\rightarrow$ 3)-Ser(Thr) unit of the glycopeptide backbone and for the disaccharide units that are commonly found in mammalian glycoconjugates. The abbreviations and codes of the lectin determinants presented in Table III will be useful for the classification of the Gal and GalNAc lectins that can be used for the study of the sugar component(s) of complex carbohydrates.

* All sugars mentioned in the tables have the D configuration (except for fucose in the L configuration), are in the pyranose form, and are linked at C-1 (except for N-acetylneuraminic acid linked at C-2), unless specifically noted.

TABLE I

Comparison of the reactivities of GalNAc-specific lectins^a

Carbohydrate residues or groups capable of reacting	Class ^b	DBL ^{20,21}	HPL ^{20,21}	AB-L ^{2,22}	WFL ^{20,23,c}	SBA ^{21,24}	LBL ^{25,26}	GSI ^{27,c} (A ₁)	VVL ²⁸ (A & B)	VVL ²⁸ (B ₁)	MPL ³⁰	BPL ³¹	SSL ^{32,c}
α GalNAc $\rightarrow$ 3GalNAc	F	3+	3+	4+	2+	d	-	d	2+	d	-	-	d
α GalNAc $\rightarrow$ 3 β GalNAc	F	4+	4+	d	3+	d	d	d	d	d	-	-	d
$\rightarrow$ 3 α Gal $\rightarrow$ 4 β Gal $\rightarrow$ 4Glc	A	+	d	3+	3+	3+	e	3+	4+	d	-	2+	d
α GalNAc $\rightarrow$ 3Gal	A	+	2+	d	3+	4+	e	3+	2+	d	-	3+	d
α GalNAc $\rightarrow$ 3 β Gal $\rightarrow$ 3GlcNAc	A	+	2+	d	3+	4+	e	3+	2+	d	-	3+	d
α GalNAc $\rightarrow$ 3(α Fuc $\rightarrow$ 2)	A ₁	2+	-	d	+	+	2+	+	2+	d	-	3+	d
β Gal $\rightarrow$ 4 β GlcNAc $\rightarrow$ 6R ^b	T	d	d	d	d	d	d	d	d	+	4+	4+	d
β Gal $\rightarrow$ 3 α GalNAc $\rightarrow$ Ser or Thr ^f	Tn	-	+	+	3+	2+	d	d	2+	4+	3+	3+	2+
α GalNAc $\rightarrow$ Ser or Thr ^g	I	-	-	d	d	+	d	d	d	d	-	3+	d
β Gal $\rightarrow$ 3GlcNAc $\rightarrow$ R ^b	II	-	-	d	+	+	d	d	d	d	-	3+	d
β Gal $\rightarrow$ 4GlcNAc $\rightarrow$ R ^b		-	-	d	+	+	d	d	d	d	-	3+	d

^aBased mainly on the results obtained from precipitin-inhibition assays: 4+, best inhibitor; 3+, about 1/2 of the best; 2+, about 1/10 of the best; +, about 1/100 of the best; -, less than 1/100 or no inhibition. ^bSee Table III for structures. ^cThe best inhibitors are α GalNAc $\rightarrow$ 6Gal and α GalNAc $\rightarrow$ 4Glc, an A active hexasaccharide²³ for WFL and GSI (A₁), respectively; α GalNAc $\rightarrow$ Ser(α GalNAc $\rightarrow$)Thr for SSL; and α GalNAc $\rightarrow$ 3(α Fuc $\rightarrow$ 2) β Gal $\rightarrow$ 3 β GlcNAc $\rightarrow$ 3 β Gal $\rightarrow$ 4Glc, an A active hexasaccharide²³ for LBL. ^dNot determined. ^eFrom the results of α GalNAc $\rightarrow$ 3(Fuc $\rightarrow$ 2) β Gal $\rightarrow$ 4GlcNAc (A active, tetra-); 3+ and α Gal $\rightarrow$ 3(α Fuc $\rightarrow$ 2)Gal, 2+, it can be concluded that Fuc is important for binding. ^fValues obtained from β Gal $\rightarrow$ 3GalNAc related inhibitors. ^gValues obtained from α GalNAc glycosides and also interact well with desialylated ovine salivary mucus glycoprotein, and mild acid treated armadillo salivary glycoprotein [GalNAc $\rightarrow$ Ser(Thr) of the protein core] (unpublished data). ^hR, related carbohydrate residues.

TABLE II

Comparison of the reactivities of β Gal $\rightarrow$ linked-specific lectins^a

Carbohydrate resi- dues or groups capa- ble of reaction	Class of lectin ^b	β Gal $\rightarrow$ 3GalNAc					β Gal $\rightarrow$ 3(4) GlcNAc					α Gal $\rightarrow$ 3Gal	
		PNA ⁴⁶	MPL ³⁰	BPL ³¹	SJL ^{33c}	RCa ₂ ³⁴	AIL ^{35d}	ALL ³⁶	RCA ₁ ³⁷	ECL ³⁸	GCL ³⁹	TAL ¹⁹	GSI-B ₄ ²⁷
β Gal $\rightarrow$ 3 α Gal- NAc $\rightarrow$ O	T	4+	4+	4+	4+	4+	4+	4+	e	e	e	e	e
α GalNAc $\rightarrow$ Ser (Thr)	Tn	-	4+	3+	?	2+	2+	2+	e	+	+	e	-
β Gal $\rightarrow$ 4 β GlcNAc $\rightarrow$ R	II	2+	-	3+	2+	2+	$\pm^d$	4+	3+	3+	4+	+ ^g	e
β Gal $\rightarrow$ 3 β GlcNAc $\rightarrow$ R	I	+	-	3+	2+	e	$\pm^d$	3+	2+	4+	4+	e	e
α Gal $\rightarrow$ 3Gal α GalNAc	B	+	-	2+	-	e	e	e	2+	+	e	e	4+
$\rightarrow$ 3 β Gal $\rightarrow$ R	A	-	-	2+	-	e	e	e	e	e	e	e	-

^a Based mainly on the results obtained from precipitin-inhibition assays: 4+, best inhibitors; 3+, about 1/2 the best; 2+, about 1/10 of the best; +, about 1/100 of the best; $\pm$, less than 1/100; -, no inhibition. ^b Representative structures shown in Table III. ^c The best inhibitor for SJL is β Gal $\rightarrow$ 3 β GalNAc-*N*-tosyl-L-serine. ^d Based on association constant³⁵. ^e Not determined. ^f The specificity for α GalNAc $\rightarrow$ 3Ser or Thr is estimated from the results of precipitin-inhibition assays; the inhibitory potency of GalNAc for PNA, that of α GalNAc-linked glycosides for MPL and BPL, and that of methyl α GalNAc for ECL. The reactivity of SJL towards this residue is questionable. ^g The best inhibitor for TAL was a biantennary saccharide with two β Gal $\rightarrow$ 4 β GlcNAc residues (see structure I) at the nonreducing end¹⁹ as 4+.

TABLE III

Classification of 2-acetamido-2-deoxy-D-galactose- and D-galactose-specific lectins

<i>Determinant</i>			
	<i>Class</i>	<i>Structure</i>	<i>Source and others</i>
1	F	$\alpha\text{GalNAc} \rightarrow 3\text{GalNAc}$	Forssman specific disaccharide
2	A	$\alpha\text{GalNAc} \rightarrow 3\text{Gal}$	Human blood group A specific disaccharide
3	A _F	$\alpha\text{GalNAc} \rightarrow 3(\alpha\text{Fuc} \rightarrow 2)\text{Gal}$	Tn antigen
	Tn	$\alpha\text{GalNAc} \rightarrow 3\text{Ser/Thr of protein core}$	
4	T	$\beta\text{Gal} \rightarrow 3\alpha\text{GalNAc} \rightarrow 3\text{Ser/Thr of protein core}$	The mucin-type sugar sequence on the human erythrocyte membrane Gangliosides
		$\beta\text{Gal} \rightarrow 3\beta\text{GalNAc} \dots \text{ceramide}$	
5	I	$\beta\text{Gal} \rightarrow 3(4)\text{GlcNAc}$	Human blood group type I and II carbohydrate sequences ⁴⁰ . Most of the lectins reactive to II are also reactive to I. Lectin I(II) determinants can be found at the nonreducing end of the carbohydrate chains derived from either <i>N</i> or <i>O</i> -glycans.
	II	$\beta\text{Gal} \rightarrow 4\text{GlcNAc}$	
6	B	$\alpha\text{Gal} \rightarrow 3\text{Gal}$	Human blood group B specific disaccharide

The source and structural relationship of the lectin determinants are shown in Scheme 1. Most of the lectin F determinants are found in glycosphingolipids⁴¹⁻⁴³, and the other determinants are present in mammalian glycoconjugates, especially in the human blood groups A, B, H, Le^a, Le^b, and Ii-active glycoproteins prepared from human ovarian cyst fluid^{44,45}. The reactivities of lectin determinants (lectin-active disaccharides) represent a combination of the binding of two individual sugars. The contribution of each sugar to the binding is not necessarily equal, and is different among lectins. For example, both *Maclura pomifera* (MPL) and peanut (PNA) lectins are $\beta\text{Gal} \rightarrow 3\text{GalNAc}$ specific^{30,46}, however, the inhibitory profile of the monosaccharide with these two lectins is quite different; $\text{GalNAc} > \text{Gal}$ in MPL and $\text{Gal} > > \text{GalNAc}$ (inactive) in PNA. From the data available in Tables I and II, a novel system to express the relationship among the carbohydrate specificities of Gal and GalNAc lectins is illustrated in Table IV.

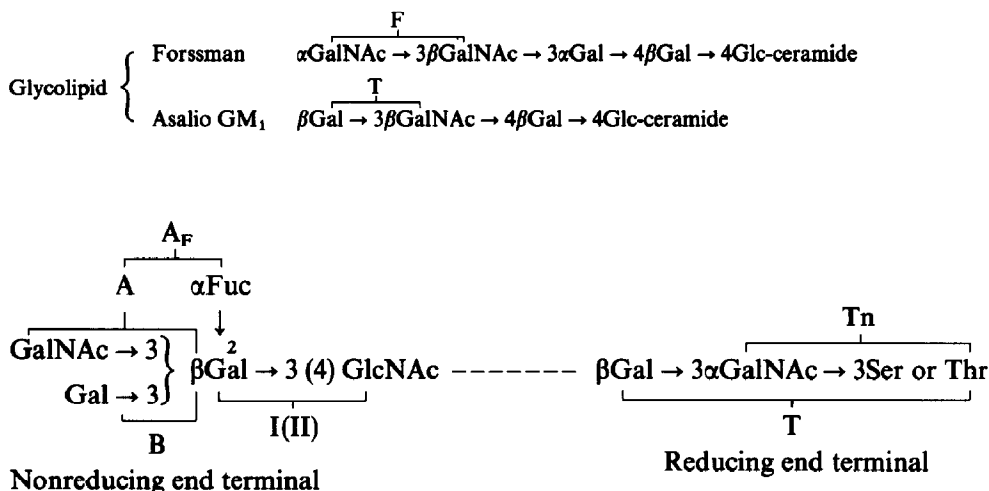
TABLE IV

Gal/GalNAc-specific lectins

<i>Class (specificity)</i>	<i>Name</i>	<i>Determinants (Active carbohydrate sequence)</i>
F/A	<i>Dolichos biflorus</i> (DBL)	F > A _r ^a > A > Tn
	<i>Helix pomatia</i> (HPL)	F > A (> A _r ^b) ≥ Tn, T
	Hog peanut (ABL, <i>Amphicarpa bracteata</i>)	F > A > Tn
	<i>Wistaria floribunda</i> (WFL)	A(A _r ^b , F > Tn, I(II))
A	Lima bean (LBL)	Hexa-A _r ^a > A _r ^b > > B
	<i>Griffonia</i> (<i>Bandeiraea</i>) <i>simplicifolia</i> -A ₄ (GSI-A ₄)	A > A _r ^b > > B
	Soy bean (SBL)	A(> A _r ^b), Tn and I(II)
	<i>Vicia villosa</i> (VVL) (A mixture of A ₄ , A ₂ B ₂ , and B ₄)	A(> A _r ^b) and Tn mainly
Tn	<i>Vicia villosa</i> B4 (VVL-B4)	Two Tn > > one Tn > > one or two T
	<i>Salvia sclarea</i> (SSL)	Two Tn > single or three sequential Tn structures
T	Peanut (<i>Arachis hypogaea</i> , PNA)	T > > I(II)
	<i>Maclura pomifera</i> (MPL)	T > Tn
	<i>Bauhinia purpurea alba</i> (BPL)	T > I(II) and Tn
	<i>Sophora japonica</i> (SJL) ^c	T and I(II)
	<i>Artocarpus integrifolia</i> (jacalin, AIL)	T > Tn > > > I(II)
	<i>Ricinus communis</i> toxin (ricin, RCA2)	T > I(II) and Tn
I(II)	<i>Ricinus communis</i> agglutinin (RCA ₁)	I(II) > T and B
	<i>Datura stramonium</i> (TAL, thorn apple)	Biantennary I(II) (Penta-2,6) > > C ^d
	<i>Erythrina cristagalli</i> (coral tree, ECL)	Multiple antennary
	<i>Geodia cydonium</i> (GLC)	I(II) and Tn
B	<i>Griffonia</i> (<i>Bandeiraea</i>) <i>simplicifolia</i> -B ₄ (GSI-B ₄)	B

^a Substitution of αFuc→2 to subterminal Gal is an important factor for binding. ^b Substitution of αFuc→2 to subterminal Gal blocks binding. ^c Some helping factor is required for precipitation of the lectin. ^d C, chitin oligosaccharide.

On this basis, the lectins determined by other methods and the lectins that require more information for classification can be included. Thus, the GalNAc-specific lectins could be subgrouped as shown in Tables V–VIII and the Gal-specific lectins in Tables IX–XII.



Scheme 1. Proposed Gal- and GalNAc-specific lectin determinants. Lectin F determinant is found in glycosphingolipid (the principal glycolipid of mammalian tissues) of the tissues of the guinea pig, horse, cat, and chicken. It can also be found on the surface of some bacteria, viruses, human gastric carcinoma, and colon tumors⁴¹⁻⁴³. All (F, A/Af, B, I/II, T, and Tn) determinants are found in human blood groups A, B, H, Le^a, Le^b, and Ii active glycoproteins, prepared from human ovarian cyst fluid^{44,45}. The $\beta\text{Gal} \rightarrow 3\beta\text{GalNAc}$ group at the nonreducing end of asialo-GM₁ is also considered as a lectin T determinant. Determinants of lectins I and II discussed in this article are equivalent to human blood group type I (Lacto-*N*-biose, $\beta\text{Gal} \rightarrow 3\text{GlcNAc}$) and type II (*N*-acetylglucosamine) carbohydrate sequences⁴⁰.

TABLE V

I-1. $\alpha\text{GalNAc} \rightarrow 3\text{GalNAc}$ (Forssman, F) and $\alpha\text{GalNAc} \rightarrow 3\text{Gal}$ (human blood group A)-specific lectins

DBL, *Dolichos biflorus* lectin (horse Gram)^{20a}.

$\alpha\text{GalNAc} \rightarrow 3\beta\text{GalNAc} \rightarrow 3\alpha\text{Gal} \rightarrow 4\beta\text{Gal} \rightarrow 4\text{Glc}$ (Forssman-specific pentasaccharide) > > $\alpha\text{GalNAc} \rightarrow 3\text{GalNAc}$ > > $\alpha\text{GalNAc} \rightarrow 3[\alpha\text{Fuc} \rightarrow 2]\beta\text{Gal} \rightarrow 4\beta\text{GlcNAc} \rightarrow 6\text{R}$ (Blood group A active pentasaccharide, R_{Lac} , 0.52°) > $\alpha\text{GalNAc} \rightarrow 3\beta\text{Gal} \rightarrow 3\text{GlcNAc}$ (A₂II)^a and $\alpha\text{GalNAc} \rightarrow 3\text{Gal}$ (R_{Lac} , 1.85°) > GalNAc.

HPL, *Helix pomatia* lectin (edible snail)^{20a,21}.

$\alpha\text{GalNAc} \rightarrow 3\beta\text{GalNAc} \rightarrow 3\alpha\text{Gal} \rightarrow 4\beta\text{Gal} \rightarrow 4\text{Glc}$ (Forssman-specific pentasaccharide) > $\alpha\text{GalNAc} \rightarrow 3\text{Gal}$ > $\alpha\text{GalNAc} \rightarrow 3\beta\text{GalNAc} \rightarrow 3\text{GlcNAc}$ (A₂II)^a > GalNAc.

ABrL, *Amphicarpaea bracteata* lectin (hog-peanut)²².

$\alpha\text{GalNAc} \rightarrow 3\text{GalNAc}$ > $\alpha\text{GalNAc} \rightarrow 3\text{Gal}$ > GalNAc.

WFL, *Wistaria floribunda* lectin^{20a,23}.

$\alpha\text{GalNAc} \rightarrow 6\text{Gal}$ > $\alpha\text{GalNAc} \rightarrow 3\text{Gal}$ (R_{Lac} , 1.85°), $\alpha\text{GalNAc} \rightarrow 3\beta\text{GalNAc} \rightarrow 3\alpha\text{Gal} \rightarrow 4\beta\text{Gal} \rightarrow 4\text{Glc}$ (Forssman-specific pentasaccharide) and $\alpha\text{GalNAc} \rightarrow 3\beta\text{Gal} \rightarrow 3\text{GlcNAc}$ (A₂II)^a > $\beta\text{GalNAc} \rightarrow 3\alpha\text{Gal} \rightarrow 4\beta\text{Gal} \rightarrow 4\text{Glc}$ (Globoside) > $\alpha\text{GalNAc} \rightarrow 3\text{GalNAc}$ > GalNAc and $\alpha\text{GalNAc} \rightarrow 3(\alpha\text{Fuc} \rightarrow 2)\beta\text{Gal} \rightarrow 4\beta\text{GlcNAc} \rightarrow 6\text{R}$ (R_{Lac} , 0.52°) > $\beta\text{Gal} \rightarrow 4\text{GlcNAc}$ (*N*-acetylglucosamine) > Lactose.

SBL, *Glycine max* lectin (soy bean) see Table VI.

^a Coded for the source or properties of oligosaccharides tested.

TABLE VI

I-2. α GalNAc $\rightarrow$ 3Gal (human blood group A)-specific lectins

GSI-A ₄ , <i>Griffonia (Bandeiraea) simplicifolia</i> A ₄ ^{20a,27} .	
α GalNAc $\rightarrow$ 3 β Gal $\rightarrow$ 3GlcNAc (A ₂ II ^a), α GalNAc $\rightarrow$ 3Gal (R_{LAC} , 1.85 ^a), and α GalNAc $\rightarrow$ 6Gal, α GalNAc $\rightarrow$ R > α Gal $\rightarrow$ 3Gal, α Gal $\rightarrow$ 6Glc, and raffinose > Gal.	
LBL, <i>Phaseolus lunatus</i> lectin (lima bean) ^{20a,21,25} .	
α GalNAc $\rightarrow$ 3(α Fuc $\rightarrow$ 2) β Gal $\rightarrow$ 3 β GlcNAc $\rightarrow$ 3 β Gal $\rightarrow$ 4Glc > α GalNAc $\rightarrow$ 3(α Fuc $\rightarrow$ 2) β Gal $\rightarrow$ 4Glc > α GalNAc $\rightarrow$ 3(α Fuc $\rightarrow$ 2) β Gal $\rightarrow$ 4 β GlcNAc $\rightarrow$ 6R (R_{LAC} , 0.56 ^a) > α GalNAc $\rightarrow$ 3(α Fuc $\rightarrow$ 2)Gal > > GalNAc > > Gal. α Fuc $\rightarrow$ 2 to subterminal Gal is an important factor for binding.	
SBL, <i>Glycine max</i> lectin (soy bean) ^{20a,24,47} .	
α - or β -GalNAc, α GalNAc $\rightarrow$ 3 β Gal $\rightarrow$ 3GlcNAc (A ₂ II ^a) and α GalNAc $\rightarrow$ 3Gal (R_{LAC} 1.85 ^a) > > α GalNAc $\rightarrow$ 3(α Fuc $\rightarrow$ 2) β Gal $\rightarrow$ 4 β GlcNAc $\rightarrow$ 6R (R_{LAC} 0.52 ^a), α Gal $\rightarrow$ 6Glc (melibiose), β Gal $\rightarrow$ 6Glc, raffinose, stachyose, β Gal $\rightarrow$ 4Glc (lactose), and α Fuc $\rightarrow$ 2 β Gal $\rightarrow$ 4 β GlcNAc $\rightarrow$ 6R (human blood H active oligosaccharide, JS, R_{LAC} 0.75 ^a) > Gal. α Fuc $\rightarrow$ 2 to subterminal Gal blocks Gal for binding.	
PT4, <i>Psophocarpus tetragonolobus</i> lectin ^{24a} .	
SN4, <i>Sambucus nigra</i> lectin ^{24b} .	
PTL, <i>Psophocarpus tetragonolobus</i> lectin ^{24a} .	
α GalNAc $\rightarrow$ 3 [α L Fuc $\rightarrow$ 2] β Gal $\rightarrow$ 4 [α L Fuc $\rightarrow$ 3] Glc (A-penta) α GalNAc $\rightarrow$ 3 [α L Fuc $\rightarrow$ 2] β Gal $\rightarrow$ 4Glc (A-tetra), and α GalNAc $\rightarrow$ 3 β Gal $\rightarrow$ 3GlcNAc > α GalNAc $\rightarrow$ 3[α L Fuc $\rightarrow$ 2]Gal (A-tri), α GalNAc $\rightarrow$ 3Gal (A-di), α Gal $\rightarrow$ 3 [α L Fuc $\rightarrow$ 2] β Gal $\rightarrow$ 4 [α L Fuc $\rightarrow$ 2] Glc (B-penta) α GalNAc $\rightarrow$ 3 β GalNAc $\rightarrow$ 3 α GalO (CH ₂) ₈ COOMe (Forssman trisaccharide), and GalNAc > α Gal $\rightarrow$ 3Gal (B-Di) > Gal.	
SNA-II, <i>Sambucus nigra</i> lectin ^{24b} .	
α GalNAc $\rightarrow$ 3Gal and α GalNAc $\rightarrow$ 6Gal > β GalNAc $\rightarrow$ 6Gal > GalNAc > di-(structure 1), tri-, -tetra-, -antennary oligosaccharides containing β Gal $\rightarrow$ 4GlcNAc at nonreducing end > β Gal $\rightarrow$ 3GalNAc (T) > > β Gal $\rightarrow$ 4GlcNAc > Gal.	
VVL, <i>Vicia villosa</i> lectin (hairy vetch; it is assumed that the lectin used to study the following determinant is a mixture of three isolectins: A ₄ -minor, B ₄ -major, and A ₂ B ₂ -minor ^{28,29}).	
α GalNAc $\rightarrow$ 3Gal (R_{LAC} , 1.85 ^a) > α GalNAc $\rightarrow$ 6Gal > α GalNAc $\rightarrow$ 3 β Gal $\rightarrow$ 3GlcNAc (A ₂ II ^a) and α GalNAc $\rightarrow$ 3GalNAc > > α GalNAc $\rightarrow$ 3(α Fuc $\rightarrow$ 2) β Gal $\rightarrow$ 4 β GlcNAc $\rightarrow$ 6R (R_{LAC} 0.52 ^a) > α Gal $\rightarrow$ 3Gal.	
WFL, <i>Wistaria floribunda</i> lectin (see Forssman-specific lectins, Table V, I-1).	
DBL, <i>Dolichos biflorus</i> lectin (see Forssman-specific lectins, Table V, I-1).	
HPL, <i>Helix pomatia</i> lectin (see Forssman-specific lectins, Table V, I-1).	

^a Coded for the source or properties of oligosaccharides tested.

TABLE VII

I-3. α GalNAc $\rightarrow$ 3Ser or Thr (Tn)-specific lectins

VVL-B ₄ , <i>Vicia villosa</i> B ₄ lectin (hairy vetch) ^{48,49} .	
A fetuin glycopeptide containing two α GalNAc $\rightarrow$ Ser(Thr) of the peptide chains (two Tn structures) > > single Tn structure > > a fetuin glycopeptide containing one or two β Gal $\rightarrow$ 3 α GalNAc $\rightarrow$ Ser(Thr) of the peptide chain(s) (T structure) > α NeuNAc $\rightarrow$ 3 β Gal $\rightarrow$ 3 α GalNAc $\rightarrow$ Ser(Thr) of the peptide chain.	
SSL, <i>Salvia sclarea</i> lectin ⁵⁰ .	
Glycopeptides containing two Tn structures > glycopeptides containing one Tn structure or glycopeptides containing three sequential Tn structures.	
MPL, <i>Machura pomifera</i> lectin.	
(see β Gal $\rightarrow$ 3GalNAc-specific lectins, Table IX, II-1).	
BPL, <i>Bauhinia purpurea alba</i> lectin.	
(see β Gal $\rightarrow$ 3GalNAc-specific lectins, Table IX, II-1).	

TABLE VIII

I-4. Other GalNAc-specific lectins requiring further characterization

APL, <i>Aegopodium podagraria</i> lectin ⁵¹ .
GalNAc > lactose > Gal > melibiose > raffinose > Ara.
BDL, <i>Bryonia dioica</i> lectin ⁵² .
GalNAc > lactose > melibiose > raffinose > stachyose.
CAL, <i>Caragana arborescens</i> lectin (pea tree) ⁵³ .
GalNAc > Gal > lactose and raffinose > stachyose > Ara > rhamnose.
CTL, <i>Clerodendron trichotomum</i> lectin ⁵⁴ .
GalNAc > lactose > melibiose > Gal.
CNL, <i>Clitocybe nebularis</i> lectin ⁵⁵ .
GalNAc and Gal.
DDL-I, <i>Dictyostelium discoideum</i> I lectin (Discoidin I) ^{56,57} .
GalNAc > D-Fuc > melibiose > lactose > Gal > L-Fuc.
EHY, L, <i>Eranthis hyemalis</i> lectin (winter aconite root tubers) ⁵⁸ .
GalNAc > lactose > Gal > melibiose > L-Fuc > Ara > raffinose.
EHe, L, <i>Euphorbia heterophylla</i> lectin ⁵⁹ .
GalNAc > lactose > Gal > melibiose > Fuc.
FFL, <i>Fomes fomentarius</i> lectin ⁵⁵ .
GalNAc > raffinose > Gal.
FJL, <i>Falcata japonica</i> lectin ^{55a} .
GalNAc.
HCL, <i>Hura crepitans</i> seed lectin ⁶⁰ .
GalNAc > Gal > melibiose and D-Fuc > raffinose.
MAL, <i>Macrotyloma axillare</i> anti-A lectin ⁶¹ .
GalNAc.
MCL, <i>Momordica charantia</i> lectin (bitter pear melon) ^{62,63} .
GalNAc > D-Fuc > Gal > raffinose and stachyose > melibiose and L-Ara.
OHL, <i>Ononis hircina</i> Jacq lectin ⁶⁴ .
GalNAc > lactose > Gal > GlcNAc > Ara.
PTL, <i>Psophocarpus tetragonolobus</i> lectin (winged bean) ^{65-67,24a} .
GalNAc > Gal and melibiose > raffinose > D-Fuc > lactose.
RPL, <i>Robinia pseudacacia</i> lectin ⁶⁸ .
GalNAc.
TML, <i>Tridacna maxima</i> lectin (Röding clam) ⁶⁹ .
GalNAc > β Gal $\rightarrow$ 6Gal, lactose and Gal > melibiose, raffinose, and stachyose.
VCL, <i>Vicia cracca</i> anti-A lectin ⁷⁰ .
GalNAc.
VAL-II & III, <i>Viscum album</i> -II & III lectins (mistletoe)
GalNAc > Gal ⁷¹ .

TABLE IX

II-1. β Gal $\rightarrow$ 3GalNAc (T)-specific lectins

PNA, <i>Arachis hypogaea</i> lectin (peanut) ^{46,72} .
β Gal $\rightarrow$ 3GalNAc > > β Gal $\rightarrow$ 4GlcNAc (<i>N</i> -acetylglucosamine) > Gal and β Gal $\rightarrow$ 3GlcNAc > > GalNAc (inactive).
BPL, <i>Bauhinia purpurea</i> alba lectin ^{31,73} .
β Gal $\rightarrow$ 3GalNAc > α GalNAc $\rightarrow$ 6Gal > GalNAc. β Gal $\rightarrow$ 3 β GlcNAc $\rightarrow$ 3 β Gal $\rightarrow$ 4Glc (lacto- <i>N</i> -tetraose) > β Gal $\rightarrow$ 3GlcNAc, β Gal $\rightarrow$ 6GlcNAc and α GalNAc $\rightarrow$ 3 β Gal $\rightarrow$ 3GlcNAc (A ₂ II*) > α GalNAc $\rightarrow$ 3Gal > β Gal $\rightarrow$ 4GlcNAc (<i>N</i> -acetylglucosamine) and β Gal $\rightarrow$ 3(α Fuc $\rightarrow$ 4) β GlcNAc $\rightarrow$ 3 β Gal $\rightarrow$ 4Glc (lacto- <i>N</i> -fucopentaose II) > β Gal $\rightarrow$ 4 β GlcNAc $\rightarrow$ 6Gal and lactose > Gal.
MPL, <i>Maclura pomifera</i> lectin (osage orange tree, hedge apple tree) ³⁰ .
β Gal $\rightarrow$ 3GalNAc > > α GalNAc $\rightarrow$ 6Gal > α Gal $\rightarrow$ 6Glc (melibiose), stachyose > GalNAc > Gal.

TABLE IX (continued)

SJL, <i>Sophora japonica</i> lectin (Japanese pagoda tree) ³³ .	
β Gal $\rightarrow$ 3 β GalNAc $\rightarrow$ N-tosyl-L-serine > β Gal $\rightarrow$ 3GlcNAc > β Gal $\rightarrow$ 3 β GlcNAc $\rightarrow$ 3 β Gal $\rightarrow$ 4Glc (lacto-N-tetraose) > β Gal $\rightarrow$ 3 β GlcNAc $\rightarrow$ 6Gal > β Gal $\rightarrow$ 4 β GlcNAc $\rightarrow$ 6Gal (blood group I Ma-active trisaccharide) and β Gal $\rightarrow$ 4GlcNAc > GalNAc > > lactose > Gal and melibiose.	
Some unknown mild acid sensitive compound near, the terminal group at the nonreducing end plays an important role in binding or precipitating this lectin.	
RCA ₂ , ricin ^{34,47,74} .	
$(\beta$ Gal $\rightarrow$ 3 α GalNAc $\rightarrow$ Ser or Thr) ₄ glycopeptide > > triantennary oligosaccharides containing β Gal $\rightarrow$ 4GlcNAc at nonreducing end > $(\beta$ Gal $\rightarrow$ 3 α GalNAc $\rightarrow$ Ser or Thr) ₂ glycopeptide with space between carbohydrate side-chains, β Gal $\rightarrow$ 3 α GalNAc $\rightarrow$ β Ser or Thr > > α GalNAc $\rightarrow$ Ser or Thr ³⁴ .	
The ability of RCA ₂ to bind to β Gal $\rightarrow$ 3GalNAc disaccharides appears to be markedly influenced by the relative locations of the disaccharides along the peptide backbone.	
Biantennary oligosaccharides containing β Gal $\rightarrow$ 4GlcNAc at nonreducing end > lactose > GalNAc > melibiose and Gal ^{34,47} ($\geq$ GalNAc, see ref. 74).	
VGL, <i>Vicia graminea</i> lectin (blood group N-specific) ^{75,76} .	
β Gal $\rightarrow$ 3 α GalNAc-linked in clusters and desialylated blood group N glycopeptide > untreated N glycopeptide and desialylated M glycopeptide.	
ABiL, <i>Agaricus bisporus</i> lectin (mushroom) ^{77,78} .	
β Gal $\rightarrow$ 3GalNAc-N-tosyl-L-serine.	
ACL, <i>Amaranthus caudatus</i> lectin ^{78a} .	
β Gal $\rightarrow$ 3 α GalNAc $\rightarrow$ Ser/Thr > > GalNAc > > Gal, β Gal $\rightarrow$ 3(4)GlcNAc (inactive).	
ARL, <i>Agropyrum repens</i> lectin (couch grass) ⁷⁹ .	
β Gal $\rightarrow$ 3GalNAc > GalNAc and chitin oligosaccharides (with type A cells).	
AIL, <i>Artocarpus integrifolia</i> lectin (jacalin) ^{80,81,81a} .	
β -Gal $\rightarrow$ 3GalNAc > α GalNAc $\rightarrow$ Ser/Thr > > > β Gal $\rightarrow$ 3GlcNAc, β Gal $\rightarrow$ 4GlcNAc, and β Gal $\rightarrow$ 4Glc.	
ALL, <i>Artocarpus lakoocha</i> lectin ^{36,82} .	
β Gal $\rightarrow$ 3GalNAc > α GalNAc $\rightarrow$ 3Ser/Thr > melibiose > raffinose > Gal.	

TABLE X

II-2. β Gal $\rightarrow$ 3(4)GlcNAc (human blood group type I and type II carbohydrate sequences; I & II)-specific lectins

AIDL-II, <i>Allo L-II</i> , <i>Allomyrina dichotoma</i> lectin II (hemolymph of a beetle) ^{82a,b} .	
β Gal $\rightarrow$ 4GlcNAc, substitution of their terminal Gal residues by α Neu5Ac $\rightarrow$ 6 will enhance their affinity to this lectin.	
RCA ₁ , <i>Ricinus communis</i> agglutinin (castor bean) ^{34,37,47,83} .	
Biantennary oligosaccharides containing β Gal $\rightarrow$ 4GlcNAc-linked units at nonreducing end > β Gal $\rightarrow$ 4 β GlcNAc $\rightarrow$ 6Gal (blood group I Ma specific) > β Gal $\rightarrow$ 4GlcNAc > β Gal $\rightarrow$ 3GlcNAc > lactose > α Gal $\rightarrow$ 3Gal > raffinose.	
RCA ₁ is also specific for β Gal $\rightarrow$ 3 α GalNAc $\rightarrow$ 3Ser (Thr). For further information, see ref. 34.	
TAL, <i>Datura stramonium</i> lectin (DSL, thorn apple) ^{47,84-87} .	
Penta-2,6 (1) > > Penta-2,4 (2) > $(\beta$ GlcNAc $\rightarrow$ 4) ₄ > $(\beta$ GlcNAc $\rightarrow$ 4) ₃ > β Gal $\rightarrow$ 4GlcNAc (N-acetylglucosamine), β GlcNAc $\rightarrow$ 4GlcNAc, and β Gal $\rightarrow$ 4 β GlcNAc $\rightarrow$ 6Man > α Man $\rightarrow$ 4 β Man $\rightarrow$ (GlcNAc) ₂ > β GlcNAc $\rightarrow$ 4Glc.	
More information is available in ref. 85.	
ECL, <i>Erythrina cristagalli</i> lectin (coral tree) ³⁸ .	
Tetra- and tri-antennary oligosaccharides containing β -Gal $\rightarrow$ 4GlcNAc-linked units at nonreducing end > β Gal $\rightarrow$ 4 β GlcNAc $\rightarrow$ 6Gal (blood group I Ma specific) > β Gal $\rightarrow$ 4GlcNAc > β Gal $\rightarrow$ 3GlcNAc > > lactose > > > β Gal $\rightarrow$ 6Gal and α Gal $\rightarrow$ 3Gal > raffinose.	
GCL, <i>Geodia cydonium</i> lectin ³⁹ .	
β Gal $\rightarrow$ 4GlcNAc, β Gal $\rightarrow$ 3GlcNAc, and lactose > GalNAc.	

TABLE X (continued)

LEL, *Lycopersicon esculentum* lectin (tomato)^{85a,b}.

In addition to $(\beta\text{GlcNAc}\rightarrow 4)_2$, $\beta\text{Gal}\rightarrow 4\beta\text{GlcNAc}\rightarrow 3\text{Gal}\rightarrow 4\beta\text{GlcNAc}\rightarrow 6\text{Gal-ol}$ sequence is also one of active oligosaccharides.

EVL, *Erythrina variegata* lectin^{85c}.

$\beta\text{Gal}\rightarrow 4\beta\text{GlcNAc}\rightarrow 3$ [$\beta\text{Gal}\rightarrow 4\beta\text{GlcNAc}\rightarrow 6$] Gal sugar sequence is the active one.

Other *Erythrina* lectins⁸⁸.

E. caffra⁺ (ECaf, L), *E. corallodendron* (ECor, L), *E. flabelliformis* (EFL), *E. humeana* (EHL), *E. latissima* (ELat, L), *E. lysistemon* (Elys, L), *E. perrierie*⁺ (EPL), *E. stricta* (ESL), and *E. zeyheri* (EZL).

Tri- or bi-antennary oligosaccharides containing $\beta\text{Gal}\rightarrow 4\text{GlcNAc}$ at nonreducing end > *N*-acetyl-lactosamine > lactose $\geq$ GalNAc > Gal (GalNAc $\geq$ lactose $\geq$ Gal).

PHA-E, *Phaseolus vulgaris* (isolectin E₄) erythroagglutinin (red kidney bean)⁸⁹⁻⁹¹.

Structure 3, in which R¹ and R² represent either hydrogen atoms or sugars, and R³ represents either GlcNAc or $\alpha\text{Fuc}\rightarrow 6$ -GlcNAc, is the minimum structural unit.

PAL, *Phytolacca americana* lectin (pokeweed mitogen).

Three different carbohydrate specificities have been reported; their relationship has not been established. $\beta\text{Gal}\rightarrow 4\beta\text{GlcNAc}\rightarrow 6$ linked^{85a}. Branched poly($\beta\text{Gal}\rightarrow 4\text{GlcNAc}$) complex⁹², ($\beta\text{GlcNAc}\rightarrow 4$)_{n>5} units⁹³, or $\alpha\text{Man}\rightarrow 2\alpha\text{Man}\rightarrow 2\alpha\text{Man}\rightarrow 3\beta\text{Man}\rightarrow 4\beta\text{GlcNAc}\rightarrow 4\text{GlcNAc-Asn}$ ⁹⁴.

PHA-L, *Phaseolus vulgaris* (isolectin L₄) leukoagglutinin^{89,95}.

Tri- and tetra-antennary oligosaccharides containing $\beta\text{Gal}\rightarrow 4\text{GlcNAc}$ -linked units at nonreducing end > > Penta-2,6 (1) > Tri-2,6 (4) > 5 > $\beta\text{Gal}\rightarrow 4\beta\text{GlcNAc}\rightarrow 2\text{Man}$ > $\beta\text{GlcNAc}\rightarrow 2\text{Man}$.

EHL, *Eranthis hyemalis* agglutinin (from winter-aconite root tubers)⁹⁶.

$\beta\text{Gal}\rightarrow 4\text{GlcNAc}$ > $\beta\text{Gal}\rightarrow 4\text{Glc}$ > Gal, melibiose > raffinose.

PNL, *Arachis hypogaea* lectin (see $\beta\text{Gal}\rightarrow 3\text{GalNAc}$ -specific lectins, Table IX, II-1).

BPL, *Bauhinia purpurea alba* lectin (see $\beta\text{Gal}\rightarrow 3\text{GalNAc}$ -specific lectin, Table IX, II-1).

Ricin (see Gal β 1 $\rightarrow$ 3GalNAc specific lectins, Table IX, II-1).

UDL, *Urtica dioica* lectin (stinging nettle)⁹⁷.

$(\beta\text{GlcNAc}\rightarrow 4)_4$ > $(\beta\text{GlcNAc}\rightarrow 4)_3$ > > $\beta\text{GlcNAc}\rightarrow 4\text{GlcNAc}$ and $\beta\text{Gal}\rightarrow 4\beta\text{GlcNAc}\rightarrow 6\text{Man}$ $\geq$ $\alpha\text{Man}\rightarrow 3\beta\text{Man}\rightarrow 4\text{GlcNAc}$ > $\beta\text{Gal}\rightarrow 4\text{GlcNAc}$ (*N*-acetyl-lactosamine) > > GlcNAc > > GalNAc.

SNL, *Sambucus nigra* lectin^{98,99}.

Bovine fetuin triantennary carbohydrate chains or porcine thyroglobulin biantennary carbohydrate chains > $\alpha\text{Neu5Ac}\rightarrow 6\beta\text{Gal}\rightarrow 4\beta\text{GlcNAc}\rightarrow 3\beta\text{Gal}\rightarrow 4\text{Glc}$ > $\alpha\text{Neu5Ac}\rightarrow 6\beta\text{Gal}\rightarrow 4\text{Glc}$ > $\alpha\text{Neu5Ac}\rightarrow 6\beta\text{Gal}\rightarrow 4\text{Glc}$ > > $\alpha\text{Neu5Ac}\rightarrow 3\beta\text{Gal}\rightarrow 4\text{Glc}$ > $\alpha\text{Neu5Ac}\rightarrow 3\beta\text{Gal}\rightarrow 3\beta\text{GlcNAc}\rightarrow 3\beta\text{Gal}\rightarrow 4\text{Glc}$ > $\beta\text{Gal}\rightarrow 4\text{GlcNAc}$ and $\beta\text{GalNAc}\rightarrow 6\text{Gal}$ > $\beta\text{Gal}\rightarrow 4\text{Glc}$ and $\alpha\text{GalNAc}\rightarrow 6\text{Gal}$ > GalNAc and $\alpha\text{GalNAc}\rightarrow 3\text{Gal}$ $\geq$ Gal > melibiose, raffinose, and stachyose.

WGA, *Triticum vulgaris* agglutinin (wheat germ)^{47,85a,85d-h}.

$\beta\text{Gal}\rightarrow 4\beta\text{GlcNAc}\rightarrow 6\text{Gal}$, ($\beta\text{Gal}\rightarrow 4$) $\beta\text{GlcNAc}\rightarrow 6\text{GalNAc}$ and $(\beta\text{GlcNAc}\rightarrow 4)_{20\text{or}3}$ > $\beta\text{Gal}\rightarrow 4\beta\text{GlcNAc}\rightarrow 3\text{Gal}$ > > > $\beta\text{Gal}\rightarrow 4\text{GlcNAc}$ (*N*-acetyl-lactosamine) > $\beta\text{Gal}\rightarrow 3\text{GalNAc}$ (inactive).

In addition to $\beta\text{GlcNAc}\rightarrow 4\text{GlcNAc}$, the $\beta\text{GlcNAc}\rightarrow 6$ sequence is the most important structure for binding. $\beta\text{GlcNAc}\rightarrow 4\text{GlcNAc}$ sequence, the lectin C determinant, can be found in mammalian *N*-linked carbohydrate chains as part of core trisaccharide and $\beta\text{Gal}\rightarrow 4\beta\text{GlcNAc}\rightarrow 6\text{Gal}$ frequently found in mammalian *O*- and *N*-linked glycoproteins.

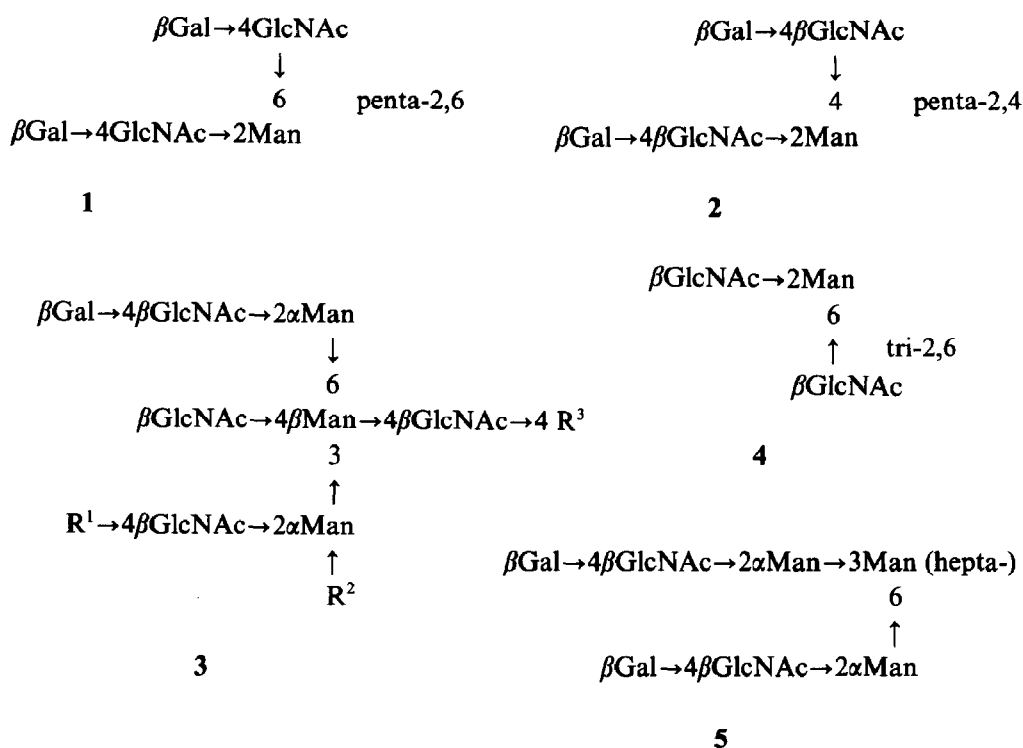


TABLE XI

II-3. $\alpha\text{Gal} \rightarrow 3\text{Gal}$ (B)-specific lectinGSL-B₄, *Griffonia (Bandeiraea) simplicifolia* I-B₄ lectin²⁷. $\alpha\text{Gal} \rightarrow 3\text{Gal} > \text{melibiose} > \text{raffinose} > \text{Gal} > > > \text{GalNAc (inactive)}$.

TABLE XII

II-4. Other Gal-specific lectins and those requiring more information for classification

AXP-I and II, *Axinella polypoides* I and II lectins¹⁰⁰. $\beta\text{Gal} \rightarrow 6\text{R} > \beta\text{Gal} \rightarrow 4\text{R} > \text{melibiose and raffinose} > \text{Gal}$.MXL, *Myxococcus xanthus* lectin¹⁰¹. $\text{NeuNAc} \rightarrow \beta\text{Gal} \rightarrow 3\text{GalNAc} \rightarrow 3\text{Ser(Thr)}$ (possibly).APL, *Abrus precatorius* lectin (jequirity bean)^{102,103}. $\text{Lactose and Gal} > \text{melibiose} > \text{Fuc}$.Abrin^{102,103}. $\text{Gal} > \text{lactose} > \text{melibiose} > \text{Fuc and Ara} > \text{Glc}$.ADL, *Aplysia depilans* lectin¹⁰⁴. $\text{Galacturonic acid} > \text{Gal}$.AML, *Ascidia malaca* lectin¹⁰⁵. $\text{Melibiose} > \text{raffinose} > \text{Gal} > \text{lactose} > \text{Ara}$.ABL, *Abramis brama* lectin¹⁰⁶. $\text{Rhamnose} > \text{Gal}$.

TABLE XII (continued)

BCL, <i>Bauhinia carronii</i> lectin ¹⁰⁷ .
Gal and lactose.
BFL, <i>Butea frondosa</i> lectin ⁶³ .
Lactose > D-Fuc > Gal > GalNAc > Ara.
CJL, <i>Crotalaria juncea</i> lectin (Sunn hemp) ¹⁰⁸ .
Lactose > melibiose > raffinose > GalNAc > Gal.
CML-I, <i>Cytisus multiflorus</i> -I lectin ¹⁰⁹ .
Gal, lactose, and melibiose > stachyose > raffinose.
CSL, <i>Cytisus scoparius</i> lectin ¹¹⁰ .
Gal and GalNAc.
DCL, <i>Didemnum candidum</i> lectin ¹¹¹ .
Lactose > Gal > melibiose > stachyose > GalNAc.
DDL-II, <i>Dictyostelium discoideum</i> -II lectin (discoidin II) ⁵⁷ .
Lactose and D-Fuc > Gal > melibiose > GalNAc > Fuc.
EAL, <i>Erythrina arborescens</i> lectin ¹¹² .
Lactose and GalNAc > melibiose and Gal > D-Fuc.
EIL, <i>Erythrina indica</i> lectin ^{63,112} .
Lactose > GalNAc and melibiose > Gal and raffinose > D-Fuc.
ELL, <i>Erythrina lithosperma</i> lectin ¹¹² .
Lactose and melibiose > GalNAc and raffinose > Gal > D-Fuc.
ESL, <i>Erythrina suberosa</i> lectin ¹¹² .
Lactose > GalNAc > melibiose and Gal > raffinose > D-Fuc.
ECL, <i>Euphorbia characias</i> L. lectin (Mediterranean spurge) ¹¹³ .
Lactose > melibiose > Gal > D-Fuc > Ara.
HRL, <i>Halocynthia roretzi</i> lectin ¹¹⁴ .
Melibiose > lactose, Gal and D-Fuc > Fuc and stachyose.
HaCL, <i>Hardenbergia comptoniana</i> lectin ¹⁰⁷ .
Raffinose.
HuCL, <i>Hura crepitans</i> latex lectin (sand-box tree) ¹¹³ .
Lactose > Gal and melibiose > Ara.
LAL, <i>Laccaria amethystina</i> lectin (LAL) (mushroom) ¹¹⁵ .
Lactose > GalNAc.
OVL, <i>Octopus vulgaris</i> lectin ¹¹⁶ .
Lactose and N-acetyllactosamine.
PAL, <i>Phaseolus aureus</i> or <i>Vigna radiata</i> lectin (mung bean) ¹¹⁷ .
Gal > Xyl > inositol.
PPL, <i>Polysphondylium pallidum</i> lectin (pallidin) ^{56,118} .
Lactose > Gal > GalNAc and D-Fuc > Fuc.
RRL, <i>Rutilus rutilus</i> lectin ¹⁰⁶ .
Rha > Gal and Ara > lactose.
SaSL, <i>Sarothamnus scoparius</i> lectin ⁵⁵ .
Raffinose > GalNAc > Gal > lactose.
SEL, <i>Scardinius erythrophthalmus</i> lectin ¹⁰⁶ .
Rha > Ara > Gal.
ViVL, <i>Vimba vimba</i> lectin ¹⁰⁶ .
Rha > raffinose > Gal and Ara > lactose.
TKL, <i>Trichosanthes kirilowii</i> lectin ¹¹⁹ .
Lactose > Gal > melibiose > raffinose.
VAL-I, <i>Viscum album</i> I lectin (mistletoe) ^{71,120} .
Lactose > melibiose, raffinose and Gal.
VUL, <i>Vigna unguiculate</i> lectin (cow-pea) ¹²¹ .
Gal.

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REFERENCES

- 1 G. L. Nicolson, *Int. Rev. Cytol.*, 39 (1974) 89–190.
- 2 I. J. Goldstein and R. D. Poretz, in I. E. Liener, N. Sharon, and I. J. Goldstein (Ed.), *The Lectins, Properties, Functions and Applications in Biology and Medicine*, Academic Press, Orlando, FL, 1986, pp. 33–247.
- 3 H. Lis and N. Sharon, in M. Sela (Ed.), *The Antigens*, Vol 4., Academic Press, New York, 1974, pp. 429–529.
- 4 H. Lis and N. Sharon, *Annu. Rev. Biochem.*, 55 (1986) 35–67.
- 5 M. E. A. Pereira and E. A. Kabat, *Crit. Rev. Immunol.*, 1 (1979) 33–78.
- 6 A. Kimura, H. Wigzell, G. Holmquist, B. Ersson, and G. Carlson, *J. Exp. Med.*, 149 (1979) 473–483.
- 7 P. Stanley, in W. J. Lennarz (Ed.), *The Biochemistry of Glycoproteins and Proteoglycans*, Plenum Press, New York, 1980, pp. 161–189.
- 8 P. Stanley, *Somatic Cell Genetics*, 9 (1983) 593–608.
- 9 W. S. Stanley, B. P. Peters, D. A. Blake, D. Yep, E. H. Y. Chu, and I. J. Goldstein, *Proc. Natl. Acad. Sci. U.S.A.*, 76 (1979) 303–307.
- 10 E. B. Briles, E. Li, and S. Kornfeld, *J. Biol. Chem.*, 252 (1977) 1107–1116.
- 11 T. Kristiansen, *Methods Enzymol.*, 34 (B) (1974) 331–341.
- 12 M. E. A. Pereira and E. A. Kabat, *J. Exp. Med.*, 143 (1976) 422–436.
- 13 W. M. Watkins, in A. Gottschalk (Ed.), *Glycoproteins, Their Composition, Structure and Function*, 2nd edn., Part B, Elsevier, Amsterdam, 1972, pp. 830–891.
- 14 R. K. Merkle and R. D. Cumming, *Methods Enzymol.*, 138 (1987) 232–259.
- 15 J. T. Gallagher, *Biosci. Rep.*, 4 (1984) 621–632.
- 16 E. A. Kabat, *Structural Concepts in Immunology and Immunochemistry*, 2nd edn, Holt, Rinehart, and Winston, New York, 1976.
- 17 O. Makela, *Ann. Med. Exp. Biol. Fenn. Suppl.*, 11 35 (1957) 1–156.
- 18 A. M. Wu and S. Sugii, *Adv. Exp. Med. Biol.*, 228 (1988) 205–263.
- 19 J. F. Crowley, I. J. Goldstein, J. Arnarp, and J. Lönngren, *Arch. Biochem. Biophys.*, 231 (1984) 524–533.
- 20 D. A. Baker, S. Sugii, E. A. Kabat, R. M. Ratcliffe, P. Hermentin, and R. U. Lemieux, *Biochemistry*, 22 (1983) 2741–2750; 20a V. Pillar, F. Piller, and J.-P. Cartron, *Eur. J. Biochem.*, 191 (1990) 461–466.
- 21 S. Hammarström, L. A. Murphy, I. J. Goldstein, and M. E. Etzler, *Biochemistry*, 16 (1977) 2750–2755.
- 22 M. J. Maliarik, D. D. Roberts, and I. J. Goldstein, *Arch. Biochem. Biophys.*, 255 (1987) 194–200.
- 23 S. Sugii and E. A. Kabat, *Biochemistry*, 19 (1980) 1192–1199.
- 24 M. E. A. Pereira, E. A. Kabat, and N. Sharon, *Carbohydr. Res.*, 37 (1974) 89–102; 24a T. Matsuda, E. A. Kabat, and A. Sürolia, *Mol. Immunol.*, 26 (1989) 189–195; 24b H. Kaku, W. J. Peumans, and I. J. Goldstein, *Arch. Biochem. Biophys.*, 277 (1990) 255–262.
- 25 S. K. Sikder, E. A. Kabat, D. D. Roberts, and I. J. Goldstein, *Carbohydr. Res.*, 151 (1986) 247–260.
- 26 D. D. Roberts and I. J. Goldstein, *J. Biol. Chem.*, 259 (1984) 903–908.
- 27 C. Wood, E. A. Kabat, L. A. Murphy, and I. J. Goldstein, *Arch. Biochem. Biophys.*, 198 (1979) 1–11.
- 28 P. M. Kaladas, E. A. Kabat, A. Kimura, and B. Ersson, *Mol. Immunol.*, 18 (1981) 969–977.
- 29 S. E. Tollerfsen and R. Kornfeld, *J. Biol. Chem.*, 258 (1983) 5172–5176.
- 30 M. Sarker, A. M. Wu, and E. A. Kabat, *Arch. Biochem. Biophys.*, 209 (1981) 204–218.
- 31 A. M. Wu, E. A. Kabat, F. G. Gruezo, and H. J. Allen, *Arch. Biochem. Biophys.*, 204 (1980) 622–639.
- 32 V. Piller, F. Piller, and J. P. Cartron, *J. Biol. Chem.*, 261 (1986) 14 069–14 075.
- 33 A. M. Wu, E. A. Kabat, F. G. Gruezo, and R. D. Poretz, *Arch. Biochem. Biophys.*, 209 (1981) 191–203.
- 34 J. U. Baenziger and D. Fiete, *J. Biol. Chem.*, 254 (1979) 9795–9799.

- 35 M. V. K. Sastry, P. Banarjee, S. R. Patanjali, M. J. Swamy, G. V. Swarnalatha, and A. Surolia, *J. Biol. Chem.*, 261 (1986) 11 726–11 733.
- 36 B. P. Chatterjee, S. Chowdhury, and H. Ahmed, *Abstr. Eur. Carbohydr. Symp.*, 4th, B-8.
- 37 A. M. Wu, S. Sugii, F. G. Gruezo, and E. A. Kabat, *Carbohydr. Res.*, 178 (1988) 243–257.
- 38 P. M. Kaladas, E. A. Kabat, J. L. Iglesias, H. Lis, and N. Sharon, *Arch. Biochem. Biophys.*, 217 (1982) 624–637.
- 39 H. Bretting, S. G. Phillips, H. J. Klumpart, and E. A. Kabat, *J. Immunol.*, 127 (1981) 1652–1658.
- 40 W. M. Watkins, *Adv. Hum. Genet.*, 10 (1980) 1–136.
- 41 G. F. Springer, *Prog. Allergy*, 15 (1971) 9–77.
- 42 A. Makita, C. Suzuki, and Z. Yosizawa, *J. Biochem. (Tokyo)*, 60 (1966) 502–513.
- 43 S. -I. Hakomori and R. Kannagi, *J. Nat. Cancer Inst.*, 71 (1983) 231–351.
- 44 A. M. Wu, *Adv. Exp. Med. Biol.*, 228 (1988) 351–394.
- 45 F. Maisonrouge-McAuliffe and E. A. Kabat, *Arch. Biochem. Biophys.*, 175 (1976) 90–113.
- 46 M. E. A. Pereira, E. A. Kabat, R. Lotan, and N. Sharon, *Carbohydr. Res.*, 51 (1976) 107–118.
- 47 H. Debray, D. Decout, G. Strecker, G. Spik, and J. Montreuil, *Eur. J. Biochem.*, 117 (1981) 41–55.
- 48 S. E. Tollefsen and R. Kornfeld, *J. Biol. Chem.*, 258 (1983) 5166–5171.
- 49 S. E. Tollefsen and R. Kornfeld, *J. Biol. Chem.*, 258 (1983) 5172–5176.
- 50 V. Piller, F. Piller, and J. P. Cartron, *J. Biol. Chem.*, 261 (1986) 14 069–14 075.
- 51 W. J. Peumans, M. Nsimba-Lubaki, B. Peeters, and W. F. Broeckaert, *Planta*, 164 (1985) 75–85.
- 52 W. J. Peumans, M. Nsimba-Lubaki, A. R. Carlier, and E. Van Driessche, *Planta*, 160 (1984) 222–228.
- 53 R. Bloch, J. Jenkins, J. Roth, and M. M. Burger, *J. Biol. Chem.*, 251 (1976) 5929–5935.
- 54 H. Kitagaki, H. Seno, H. Yamaguchi, and I. Matsumoto, *J. Biochem. (Tokyo)*, 97 (1985) 791–799.
- 55 V. Hořejší and J. Kocourek, *Biochim. Biophys. Acta*, 538 (1978) 299–315; 55a T. Nakajima, S. Yazawa, T. Kogure, and K. Kurukawa, *Biochem. Biophys. Acta*, 964 (1988) 207–212.
- 56 S. D. Rosen, D. L. Simpson, J. E. Rose, and S. H. Barondes, *Nature (London)*, 252 (1974) 128 and 149–151.
- 57 W. A. Frazier, S. D. Rosen, R. W. Reitherman, and S. H. Barondes, *J. Biol. Chem.*, 250 (1975) 7714–7721.
- 58 B. R. Cammue, B. Peeters, and W. J. Peumans, *Biochem. J.*, 227 (1985) 949–955.
- 59 M. Nsimba-Lubaki, W. J. Peumans, and A. R. Carlier, *Biochem. J.*, 215 (1983) 141–145.
- 60 A. Falasca, C. Franceschi, C. A. Rossi, and F. Stirpe, *Biochim. Biophys. Acta*, 602 (1980) 95–105.
- 61 T. Haylett and L. S. Swart, *S. Afr. J. Chem.*, 35 (1982) 33–36.
- 62 T. Mazumder, N. Gaur, and A. Surolia, *Eur. J. Biochem.*, 113 (1981) 463–470.
- 63 V. Hořejší, M. Ticha, J. Novotny, and J. Kocourek, *Biochim. Biophys. Acta*, 623 (1980) 439–448.
- 64 V. Hořejší, O. Chaloupecka, and J. Kocourek, *Biochim. Biophys. Acta*, 538 (1979) 287–293.
- 65 P. S. Appukutan and D. Basu, *Anal. Biochem.*, 113 (1981) 253–255.
- 66 A. A. Kortt, *Eur. J. Biochem.*, 138 (1984) 519–525.
- 67 A. A. Kortt, *Arch. Biochem. Biophys.*, 236 (1985) 544–554.
- 68 V. Hořejší, C. Haskovec, and J. Kocourek, *Biochim. Biophys. Acta*, 532 (1978) 98–104.
- 69 B. A. Baldo, W. H. Sawyer, R. V. Stick, and G. Uhlenbruck, *Biochem. J.*, 175 (1978) 467–477.
- 70 H. Rüdiger, *Eur. J. Biochem.*, 72 (1977) 317–322.
- 71 H. Franz, P. Ziska, and A. Kindt, *Biochem. J.*, 195 (1981) 481–484.
- 72 R. Lotan and N. Sharon, *Methods Enzymol.*, 50 (1978) 361–367.
- 73 T. Osawa, T. Irimura, and T. Kawaguchi, *Methods Enzymol.*, 50 (1978) 367–372.
- 74 G. L. Nicolson, J. Blaustein, and M. E. Etzler, *Biochemistry*, 13 (1974) 196–204.
- 75 M. Duk and E. Lisowska, *Eur. J. Biochem.*, 118 (1981) 131–136.
- 76 M. Duk, E. Lisowska, M. Kordowicz, and K. Warniowska, *Eur. J. Biochem.*, 123 (1982) 105–112.
- 77 C. A. Presant and S. Kornfeld, *J. Biol. Chem.*, 247 (1972) 6937–6945.
- 78 S. Sueyoshi, T. Tsuji, and T. Osawa, *Biol. Chem. Hoppe-Seyler*, 366 (1985) 213–221; 78a S. J. Rinderle, I. J. Goldstein, K. L. Matta, and R. M. Ratcliffe, *J. Biol. Chem.*, 264 (1989) 16123–16131.
- 79 B. Cammue, H. M. Stinissen, and W. J. Peumans, *Eur. J. Biochem.*, 148 (1985) 315–322.
- 80 M. V. K. Sastry, P. Banarjee, S. R. Patanjali, M. J. Swamy, G. V. Swarnalatha, and A. Surolia, *J. Biol. Chem.*, 261 (1986) 11 726–11 733.
- 81 H. Ahmed and B. P. Chatterjee, *J. Biol. Chem.*, 264 (1989) 9365–9372; 81a S. K. Mahanta, K. Sastry and A. Surolia, *Biochem. J.*, 265 (1990) 831–840.

- 82 B. P. Chatterjee, H. Ahmed, S. Chowdhury, and S. Ray, *Int. Symp. Lectins*, Israel, Sept. 17–19, 1989; 82a K. Yamashita, K. Umetsu, T. Suzuki, Y. Iwaki, T. Endo, and A. Kobata, *J. Biol. Chem.*, 263 (1988) 17482–17489; 82b S. Süeyoshi, K. Yamamoto, and T. Osawa, *J. Biochem.*, 103 (1988) 894–899.
- 83 R. Kaifu and T. Osawa, *Carbohydr. Res.*, 52 (1976) 179–185.
- 84 N. N. Desai, A. K. Allen, and A. Neuberger, *Biochem. J.*, 197 (1981) 345–353.
- 85 J. F. Crowley, I. J. Goldstein, J. Arnarp, and J. Lönnngren, *Arch. Biochem. Biophys.*, 231 (1984) 524–533; 85a H. Kawahima, S. Süeyoshi, H. Li, K. Yamamoto, and T. Osawa, *Glycoconjugate J.* 7 (1990) 323–334; 85b B. C. R. Zhu, and R. A. Laine, *Eur. J. Biochem.*, 188 (1990) 67–71; 85c H. Li, K. Yamamoto, H. Kawashima, and T. Osawa, *Glycoconjugate J.*, 7 (1990) 311–322; 85d O. Renkonen, P. Makinen, K. Hard, J. Helin, L. Penttilä, *Can. J. Biochem. Cell Biol.*, 66 (1988) 449–453; 85e O. Renkonen, L. Penttilä, R. Neimelä, A. Vainio, A. Leppänen, J. Helin, A. Seppo, A. Makkonen, and H. Maaheimo, *Carbohydr. Res.*, 213 (1991) 169–182; 85f A. M. Wu, and S. Sugii and A. Herp, *Adv. Exp. Med. Biol.*, 228 (1988) 817–853; 85g I. J. Goldstein, S. Hammanström, and G. Sundblad, *Biochim. Biophys. Acta*, 404 (1975) 63–67; 85h A. K. Allen, A. Neuberger, and N. Sharon, *Biochem. J.*, 131 (1973) 155–162.
- 86 K. Yamashita, K. Totani, T. Ohkura, S. Takasaki, I. J. Goldstein, and A. Kobata, *J. Biol. Chem.*, 262 (1987) 1602–1607.
- 87 R. D. Cummings and S. Kornfeld, *J. Biol. Chem.*, 259 (1984) 6253–6260.
- 88 H. Lis, F. J. Joubert, and N. Sharon, *Phytochemistry*, 24 (1985) 2803–2809.
- 89 R. D. Cummings and S. Kornfeld, *J. Biol. Chem.*, 257 (1982) 11 230–11 234.
- 90 T. Irimura, T. Tsuji, S. Tagami, K. Yamamoto, and T. Osawa, *Biochemistry*, 20 (1981) 560–566.
- 91 K. Yamashita, A. Hitoi, and A. Kobata, *J. Biol. Chem.*, 258 (1983) 14 753–14 755.
- 92 T. Irimura and G. L. Nicolson, *Carbohydr. Res.*, 120 (1983) 187–195.
- 93 K. Yokoyama, T. Terao, and T. Osawa, *Biochim. Biophys. Acta*, 538 (1978) 384–396.
- 94 Y. Katagiri, K. Yamamoto, T. Tsuji, and T. Osawa, *Carbohydr. Res.*, 120 (1983) 283–292.
- 95 S. Hammarström, M. L. Hammarström, G. Sundblad, J. Arnarp, and J. Lönnngren, *Proc. Natl. Acad. Sci. U.S.A.*, 79 (1982) 1611–1615.
- 96 B. P. Cammue, B. Peeters, and W. J. Peumans, *Biochem. J.*, 227 (1985) 949–955.
- 97 N. Shibuya, I. J. Goldstein, J. A. Shafer, W. J. Peumans, and W. F. Broekaert, *Arch. Biochem. Biophys.*, 249 (1986) 215–224.
- 98 W. F. Broekaert, M. Nsimba-Lubaki, B. Peeters, and W. J. Peumans, *Biochem. J.*, 221 (1984) 163–169.
- 99 N. Shibuya, I. J. Goldstein, W. F. Broekaert, M. Nsimba-Lubaki, B. Peeters, and W. J. Peumans, *J. Biol. Chem.*, 262 (1987) 1596–1601.
- 100 H. Bretting and E. A. Kabat, *Biochemistry*, 15 (1976) 3228–3236.
- 101 M. Cumsy and D. R. Zusman, *Proc. Natl. Acad. Sci. U.S.A.*, 76 (1979) 5505–5509.
- 102 S. Olsnes, E. Saltvedt, and A. Pihl, *J. Biol. Chem.*, 249 (1974) 803–810.
- 103 S. Olsnes, *Methods Enzymol.*, 50 (1978) 323–330.
- 104 N. Gilboa-Garber, A. J. Susswein, L. Mizrahi, and D. Avichezer, *FEBS Lett.*, 181 (1985) 267–270.
- 105 N. Parrinello and C. Canicatti, *Dev. Comp. Immunol.*, 6 (1982) 53–64.
- 106 A. Krajhanzl, V. Hořejší, and J. Kocourek, *Biochim. Biophys. Acta*, 532 (1978) 215–224.
- 107 R. L. P. Flower, G. E. Wilcox, V. Chugg, and J. R. Neal, *Aust. J. Exp. Med. Sci.*, 62 (1984) 763–769.
- 108 B. Ersson, K. Aspberg, and J. Porath, *Biochim. Biophys. Acta*, 310 (1973) 446–452.
- 109 Y. Konami, K. Yamamoto, T. Tsuji, I. Matsumoto, and T. Osawa, *J. Pharmacobiodyn.*, 6 (1983) 737–747.
- 110 N. M. Young, D. C. Watson, and R. E. Williams, *Biochem. J.*, 222 (1984) 41–48.
- 111 G. R. Vasta and J. J. Marchalonis, *J. Biol. Chem.* 261 (1986) 9182–9186.
- 112 L. Bhattacharyya, P. K. Das, and A. Sen, *Arch. Biochem. Biophys.*, 211 (1981) 459–470.
- 113 L. Barbieri, A. Falasca, C. Franceschi, F. Licastro, C. A. Rossi, and F. Stirpe, *Biochem. J.*, 215 (1983) 433–439.
- 114 H. Yokosawa, H. Sawada, Y. Abe, T. Numakunai, and S. -L. Ishii, *Biochem. Biophys. Res. Commun.*, 107 (1982) 451–457.
- 115 J. Guillot, L. Genaud, J. Gueugnot, and M. Damez, *Biochemistry*, 22 (1983) 5365–5369.
- 116 W. Rögener, L. Renwrantz, and G. Uhlenbruck, *Dev. Comp. Immunol.*, 9 (1985) 605–616.
- 117 C. N. Hankins, and L. M. Shannon, *J. Biol. Chem.*, 253 (1978) 7791–7797.
- 118 S. D. Rosen, J. Kaur, D. L. Clark, B. T. Pardos, and W. A. Frazier, *J. Biol. Chem.*, 254 (1979) 9408–9415.

- 119 H. W. Yeung, T. B. Ng, D. M. Wong, and W. W. Li, *Int. J. Pep. Protein. Res.*, 27 (1986) 208–220.
- 120 P. Ziska and H. Franz, *Experimentia*, 37 (1981) 219.
- 121 B. J. Roberson and D. R. Strength, *Prep. Biochem.*, 13 (1983) 45–56.
- 122 H. Ahmed and B. P. Chatterjee, *Carbohydr. Res.*, 177 (1988) 173–183.